



POPULAR ARTICLE

Synthetic microbial communities (SynComs): Designing the next generation of biofertilizers and biocontrol agents

M. Tejashree¹, S. Triveni¹, J. Sai Prasad¹, P. Samata², D. Sridevi³

1 Department of Agricultural Microbiology and Bioenergy, College of Agriculture, PJTAU, Rajendranagar, Hyderabad – 500036, Telangana, India

2 Department of Agronomy, Regional Sugarcane and Rice Research Station, Rudrur, PJTAU, Nizamabad District – 503188, Telangana, India

3 Department of Agricultural Microbiology and Bioenergy, College of Agriculture, PJTAU, Rajendranagar, Hyderabad – 500036, Telangana, India

Corresponding authors email: tjsri5@gmail.com

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Intricacies of the subject

For nearly a century, agricultural microbiology has relied on single-strain bioinoculants a *Rhizobium* culture for legumes, an *Azotobacter* or *Azospirillum* formulation for cereals, a *Trichoderma* or *Pseudomonas* product for disease suppression. While these products have delivered real benefits, their field performance is notoriously inconsistent: a strain that excels in the laboratory often fails to establish against the diverse native soil microflora, or provides a function that is not the factor limiting crop growth at that site. Synthetic Microbial Communities, popularly called SynComs, represent a fundamental shift in thinking. Instead of asking “which single strain is best?”, SynCom research asks “which combination of microorganisms, assembled rationally, can reproduce the benefits of a healthy plant microbiome?” A SynCom is a defined, reduced-complexity consortium typically 3 to 25 well-characterized strains designed on ecological and functional principles so that the members complement

one another, occupy different niches, and persist together in the rhizosphere or inside plant tissues.

Why single strains fall short

The rhizosphere of a single crop plant harbors thousands of bacterial and fungal taxa engaged in competition, cross-feeding, signaling, and antagonism. When a lone inoculant is introduced into this arena, three problems commonly arise:

Poor colonization and persistence : the introduced strain is outcompeted by adapted native microflora within days or weeks.

Functional narrowness : A phosphate solubilizer does nothing for nitrogen limitation; a siderophore producer cannot suppress a pathogen that is not iron-limited.

Context dependence: performance varies with soil type, pH, moisture, crop genotype and season.

Multi-functional consortia address all three limitations simultaneously: complementary strains cover multiple growth-limiting



functions, occupy distinct micro-niches (root tip, root hair zone, endosphere), and buffer one another against environmental fluctuation.

How SynComs are designed

What distinguishes a SynCom from an ordinary “mixed biofertilizer” is rational design. The modern workflow proceeds in stages:

1. **Microbiome mining:** 16S rRNA/ITS amplicon sequencing and metagenomics of healthy, high-performing plants identify the “core microbiome” taxa consistently associated with the crop across locations and seasons — as candidate members.
2. **Culturomics:** Large-scale isolation campaigns using diverse media, dilute nutrient conditions and long incubations recover culturable representatives of the core taxa, bringing much of the “unculturable majority” into culture.
3. **Functional characterization:** Each isolate is screened for nitrogen fixation, P/K/Zn solubilization, IAA and ACC-deaminase production, siderophores, antifungal metabolites, biofilm formation and induced systemic resistance (ISR) elicitation. Whole-genome sequencing confirms these traits and rules out virulence or antibiotic-resistance genes.
4. **Compatibility and assembly:** Cross-streaking and co-culture assays eliminate mutually antagonistic combinations, while genome-scale metabolic modelling and machine-learning tools predict which strains will be metabolically complementary (cross-feeding).
5. **Iterative testing:** Candidate SynComs are validated in gnotobiotic systems, pots and field microplots, with “drop-out” experiments revealing the essential keystone members.

Recent advances driving the field

- **Reduced-complexity model communities:** Landmark studies in *Arabidopsis*, maize, rice and tomato show that SynComs of 7–15 members can reproduce much of the growth promotion and disease suppression of the full native microbiome.
- **Machine learning-guided assembly:** AI models now predict community stability and plant-beneficial outcomes from strain genomes alone, shortening the trial-and-error phase of design.
- **Stress-tailored SynComs:** Consortia combining ACC-deaminase producers, exopolysaccharide-secreting osmotolerant strains and biofilm formers show promise against drought and salinity highly relevant for rainfed and salt-affected soils of the Deccan region.
- **Endophytic and disease-suppressive SynComs:** Seed- and root-endophyte communities escape soil competition by colonizing internal tissues, while consortia pairing antibiosis-based antagonists (e.g., lipopeptide-producing *Bacillus spp.*) with ISR-eliciting strains and competitive root colonizers suppress wilt, root rot and blight complexes more consistently than any single agent a “multiple-lines-of-defence” strategy pathogens find hard to overcome.

Formulation and delivery: The practical frontier

For Indian conditions, the success of SynComs will hinge on formulation science. Carrier-based (talc, lignite) and liquid formulations must now keep several organisms viable, in balanced proportions, through storage and field application. Promising approaches include:

- Encapsulated formulations alginate or starch beads that protect each member and release them together in the spermosphere;



- Biofilm-based inoculants members co-cultured as a mixed biofilm on the carrier, arriving in the soil already organized as a community;
- Seed coating/priming and staggered delivery placing the community where the emerging root will pass, or combining seed treatment (endophytes) with soil application (rhizosphere strains) and foliar sprays.

Shelf-life, member-ratio stability and quality-control protocols (strain-specific qPCR markers to verify each member's count) are active research needs where Indian laboratories with strong biofertilizer production experience can contribute immediately.

Regulatory and adoption challenges

The Fertilizer Control Order (FCO) currently specifies standards for individual biofertilizer types (Rhizobium, Azotobacter, PSB, KSB, etc.) and a few two- or three-organism consortia. Multi-strain SynComs will require updated registration frameworks specifying identity, minimum counts per member and absence of contaminants, alongside biosafety

assessment and farmer education on storage and application. The direction of travel is nonetheless clear: defined multi-strain products have already reached international markets, and Indian public-sector institutions are well placed to develop indigenous SynComs tailored to our soils, crops and climate.

Conclusion

SynComs mark the transition of agricultural microbiology from single “silver-bullet” strains to microbiome engineering assembling teams of microbes the way an agronomist assembles a cropping system. By combining microbiome sequencing, culturomics, functional screening and computational design, researchers can now build consortia that colonize better, perform multiple functions and deliver more reproducible benefits in farmers' fields. For a country seeking to reduce chemical fertilizer and pesticide dependence while sustaining productivity, rationally designed microbial communities may become one of the defining agricultural technologies of the coming decade.

References

1. Vorholt, J.A., Vogel, C., Carlström, C.I. and Müller, D.B. 2017. Establishing causality: opportunities of synthetic communities for plant microbiome research. *Cell Host Microbe*, 22(2): 142–155.
2. de Souza, R.S.C., Armanhi, J.S.L. and Arruda, P. 2020. From microbiome to traits: designing synthetic microbial communities for improved crop resiliency. *Front. Plant Sci.*, 11: 1179.
3. Shayanthan, A., Ordoñez, P.A.C. and Oresnik, I.J. 2022. The role of synthetic microbial communities (SynCom) in sustainable agriculture. *Front. Agron.*, 4: 896307.
4. Martins, S.J. et al. 2023. Reproducible plant microbiome engineering through synthetic communities. *Trends Microbiol.*